**Team3ispA: Miniprep**

**source**: mini shaker

**samples**

*culture label side\_label destination plasmid*

p1-1 m1-1 sF\_ispA1-1 Box\_ Team3ispA/A7 pLYC73S-ispA1-1

p2-1 m2-1 yP\_ispA1-1 Box\_ Team3ispA/B7 pLYC73S-ispA2-1

p3-1 m3-1 sM\_ispA1-1 Box\_ Team3ispA/C7 pLYC73S-ispA3-1

p4-1 m4-1 cB\_ispA1-1 Box\_ Team3ispA/D7 pLYC73S-ispA4-1

p5-1 m5-1 pA\_ispA1-1 Box\_ Team3ispA/E7 pLYC73S-ispA5-1

**Protocol**

* Take an image of the culture block, upload it later to the Github issue
* For each sample, perform 2 minipreps (each on 2mL culture)
* Combine the two identical eluted minipreps in a regular zymo cleanup;

elute with **20uL** ddH2O

**Team3ispA: Sequence**

**source**:

seqispA1-F benchtop/lyophilized

**samples**

*label location plasmid oligo*

m1-1 Box\_ ispA3/A7 pLYC73S-ispA1 seqispA1-F

m2-1 Box\_ ispA3/B7 pLYC73S-ispA2 seqispA1-F

m3-1 Box\_ ispA3/C7 pLYC73S-ispA3 seqispA1-F

m4-1 Box\_ ispA3/D7 pLYC73S-ispA4 seqispA1-F

m5-1 Box\_ ispA3/E7 pLYC73S-ispA5 seqispA1-F

**dilutions**

*label concentration destination*

seqispA1-F 100uM Box\_ ispA3/G1

2.66uM seqispA1-F 2.66uM Box\_ ispA3/G2

**protocol**

* Make 100uM oligo stocks:
  + Rehydrate according to IDT sheet
* Make 2.66uM oligo stocks:
  + 97.3 uL ddH2O
  + 2.7 uL oligo
* For each plasmid listed, mix the following sequencing reactions in an eppendorf tube:
  + 4 uL ddH2O
  + 6 uL miniprep DNA (undiluted)
  + 3 uL oligo (2.66 uM)
* Clearly label the tops of the tubes with the “label”
* Take the sequencing reactions and order form to:
  + 237 Stanley Hall (second floor cold room)